Enantioselective Epoxidation of Styrene Derivatives by Chloroperoxidase Catalysis

Stefano COLONNA,^{2,4} Nicoletta GAGGERO,² Luigi CASELLA,^b Giacomo CARREA,^c and Piero PASTA^C

^aDipartimento di Chimica Organica e Industriale, Centro C.N.R., via Golgi 19, Milano, Italy; ^bDipartimento di Chimica Generale, Universita' di Pavia, via Taramelli 12, Pavia, Italy and ^CIstituto di Chimica degli Ormoni, C.N.R., via Mario Bianco 9, Milano, Italy

(Received 1 February 1993)

ABSTRACT. Chloroperoxidase catalysed epoxidation of styrene derivatives by t-BuOOH preferentially gives (R) oxides with ee values between 28 and 68%. The data support the view of oxygen delivery from the ferryl oxygen directly to the substrate.

INTRODUCTION

Asymmetric epoxidation is of fundamental importance in biological processes such as the metabolism of xenobiotics having aromatic or olefinic double bonds and the cyclisation of squalene to triterpenoids and steroids. However, in contrast with chemical reagents such as chiral (salen) manganese complex derivatives¹ or chirally modified metalloporphyrins,² heme enzymes such as cytochrome c peroxidase,³ afford epoxides with very low ee, whereas horseradish peroxidase is unable to catalyze the epoxidation of styrene.⁴ Substantially higher enantioselectivities are observed in the oxidation of styrene, and cis and trans-B-methyl styrene to the corresponding oxides⁵ with cytochrome P-450_{cam}. Even simple aliphatic alkenes are epoxidized with medium ee by this enzyme .⁶ Polycyclic aromatic epoxides are also produced with high enantioselectivity by cytochrome P-450 isozymes.⁷ Just one case has been reported of stereoselective epoxidation of a simple, unfunctionalized alkene catalyzed by the fungal enzyme chloroperoxidase (E.C. 1.11.1.10) chloride/hydrogen peroxide oxidoreductase) from Caldariomyces fumago, namely, the epoxidation of trans-[1-2H] styrene, that proceeds without detectable loss of stereochemistry.⁴ Chloroperoxidase, like cytochrome c peroxidase is an anomalous peroxidase in that it catalyzes P-450 like monooxygenation reactions. We wanted to ascertain if this enzyme is able to transfer enantioselectively an oxygen atom to the double bond and to investigate the steric course of the reaction.

RESULTS AND DISCUSSION

Preliminary results with p-chlorostyrene at pH 6 in a phosphate buffer solution at 25° C, indicated that t-BuOOH is the oxidant of choice with respect to H_2O_2 in view of the higher chemical yield obtained, whereas the ee were practically the same (66-67%) with both oxidizing agents. The time course of p-chlorostyrene oxidation by t-BuOOH and H_2O_2 is shown in Table I.

Besides epaxide, the corresponding arylacetoaldehyde and diol were concomitantly produced. Without the enzyme, the olefin substrate was not oxygenated by the oxidants alone under identical reaction conditions.

Table I. Time course of chloroperoxidase catalysed oxidation of p-chlorostyrene as a function of the nature of the oxidant.⁸

^a p-Chlorostyrene (7.2 µmol) was added to 1 ml of 0.05 M potassium phosphate buffer, pH 6, containing the oxidant $(15 \mu \text{mol})$ and chloroperoxidase (29 units) . The mixture was shaken for the scheduled time, at 25° C, diluted with acetone (2 ml) and analyzed by chiral GLC.

In order to verify the generality of the reaction, we investigated the t -BuOOH oxidation of other substituted styrenes. Chemical yield, absolute configuration and ee of the obtained epoxides are reported in Table II. The data show that in the case of chloroderivatives the para and meta substitution led to higher chemical yield than the *ortho*-substitution, without substantially affecting the enantioselectivity. This behaviour is in contrast with the results observed by us in the chloroperoxidase catalyzed sulfoxidation reaction, where p-substituted phenyl alkyl sulfides were oxidized in much higher enantioselectivity than o -substituted phenyl alkyl sulfides.⁸ On the other hand, the epoxidation of p-nitrostyrene occured in poor chemical and optical yield. The ee of the epoxides did not change for the entire reaction period. The enxymatic asymmetric synthesis was not accompanied by kinetic resolution of the product since racemic p-chlorostyrene oxide was recovered unchanged under the usual reaction conditions.

Table II. Chemical yield, ee and absolute configuration of the epoxides obtained **by** chloroperoxidase catalysed oxidation of some arylalkenes by *t*-BuOOH.^a

*** For conditions see the Experimental section. b Gn the bssis of the elution order on GLC.**

The absolute configurations of the prevailing enantiomers of styrene epoxide (R) and diol (S) **were determined by comparison with the authentic samples (Aldrich). In the case of p-chloro, m-chloro and p-bromostyrene epoxides the absolute configurations were assigned by comparison of** their circular dichroism and ¹H NMR spectra, in the presence of Eu(hfc)₃ chiral shift reagent, with those of (R) styrene oxide. In the case of o -chloro and p -nitrostyrene epoxides the absolute configurations were deduced by the elution order on chiral GLC, the (R) enantiomer being eluted before the (S) enantiomer for all the epoxides reported in Table II. The (R) absolute configuration **indicates that in all cases the approach by the oxidant to the re face of the double bond is the** preferred one. The (S) configuration of 1-phenyl-1,2-ethanediol is a consequence of a S_N2 **nucleophilic attack of Hz0 on the benzylic carbon of (R) styrene oxide formed in the course of the enzymatic reaction, without appreciable racemisation (ee of diol, 48%). This stereochemical course was confirmed by performing the hydrolysis of a specimen of the (R) oxide under the usual reaction conditions in the absence of the enzyme.**

Phenyl acetaldehydes produced as coproducts do not derive from the decomposition of the corresponding styrene oxides, as shown by Ortiz de Montellano⁴ in the case of *trans* [1-²H] styrene. **These aldehydes probably derive from an hydrogen migration within an intemrediate formed by reaction of the olefin with the oxygen-iron complex. This is the case also in the reaction catalysed** by cytochrome P-450⁹ or by chiral iron porphyrins.¹⁰

The influence of the addition of water-miscible cosolvents on the stereoselectivity of the oxidation of p-chloro styrene with t-BuOOH was also examined. Dioxane and acetone decreased **both the optical and the chemical yield of the epoxide; for instance, at 10% concentration (v/v) ee** values were 48% and 44%, and the rates were 30% and 40% lower than in buffer alone. On the other hand, acetonitrile only slightly affected the enantioselectiviy but, as a function of its **concentration, modified the products ratio by favouring the formation of p-chlorophenyl** acetaldehyde and diol at the expenses of the epoxide (Table III).

Table III. Influence of acetonitrile concentration (v/v) on chloroperoxidase catalyzed oxidation of **p-chlorostyrene by ~-BuOOH.~**

a For **conditions see the legend to Table I.**

The use of magnesium monoperoxyphtalate, an efficient oxygen donor for manganese porphyrin catalyzed epoxidation of alkenes,¹¹ gave unsatisfactory results. Indeed, starting from **p-chlorostymne the corresponding epoxide was obtained in 30% chemical yield and 30% cc,** together with p-chlorophenyl acetaldehyde (10%), for a 300 min reaction time.

It is assumed that the mechanism of the epoxidation of olefin by chloroperoxidase⁴ and cytochrome c peroxidase³ with H_2O_2 involves a ferryl oxygen transfer from compound I. The substantial enantioselectivity observed by us for the first time in the chloroperoxidase oxidation of styrene derivatives with t -BuOOH (and H₂O₂) supports the view of oxygen delivery from the ferryl oxygen directly to the substrate. This also indicates that with the former enzyme the ferry1 oxygen is more accessible to the olefins than with the latter **one** which gives low mandoselectivity.3 **As** a matter of fact, in spite of the different steric and electronic requirements of the sulfoxidation and epoxidation reactions, chloroperoxidase affords almost enantiomerically pure sulfoxides⁸ also in the oxidation of sulfides, whereas cytochrome c peroxidase leads to nearly racemic products.⁴ Finally, it is worth **mentioning** that an iron-oxo **intermcdiatc** has been postulated by Groves in the porphyrin-catalysed epoxidation of styrene¹⁰ that leads preferentially, as in our case, to the (R) enantiomer of styrene oxide.

EXPERIMENTAL SECTION

Materials. Styrene, *o*-chlorostyrene, *m*-chlorostyrene, *p*-chlorostyrene and *p*-bromostyrene were commercial products from Aldrich. p -Nitrostyrene was prepared according to the literature.¹²

Styrene, o, m, and p-chlorostyrene, p-bromostyrene and p-nitrostyrene epoxides are known in the optically active form.^{2,10,13} Chloroperoxidase from *Caldariomyces fumago* [RZ 0.6] was obtained from Sigma.

General Methods. The optical rotations were determined with a Perkin Elmer R241 polarimeter. The ¹H NMR spectra of the products were recorded in CDC1 $_3$ on a Varian 390 instrument. The CD spectra were recorded on a Jasco J-500 dichrograph.

Enzymatic Oxidations: typical procedure. The alkene (1 mmol) and CFG (1000 units) were magnetically stirred in 140 mL of 0.05 M phosphate buffer, pH 6, at 25°C. The oxidant (2 mmol) in 7.5 mL of buffer solution pH 6, was added in 150 min in 30 aliquots at 5 min intervals and the reaction was carried out for 24 h. **Extraction** with 2 portions (200 mL each) of diethyl ether, followed by drying and evaporation of the organic solvent, gave the crude product, that was purified by column chromatography on florisil with mixtures of diethyl ether and light petroleum.

Determination of degree of conversion and enantiomeric excess, Both the degree of conversion of substrates into epoxides, aldehydes and dials, and the ee values of epoxides and diols were determined, in one run, by chiral GLC with a CP-Cyclodextrin-8-2,3,6-M19 column (50 m, 0.25 mm ID, Chrompack) under the following conditions: oven temperature 110° C (initial time 20 min) to 180°C with a heating rate of 1°C/min; H_2 as carrier gas. Epoxide and diol enantiomers were base line separated.

REFERENCES

- (1) E.N. Jacobsen, W. Zhang, A.R. Muci, J.R. Ecker, C. Deng J. Amer. Chem. Soc., 1991, 113, 7063 and references therein.
- (2) Y. Naruta. F. Tani, N. Ishihara. K. Maruyarna J. Amer. Chem. Sot., **l!Bl.Z13,6865 and references therein.**
- **(3) V.P. Miller, G.D. De Pillis, J.C. Ferrer, A.** Grant Mauk, **P.R. ortiz de Montellano J.** *Biol. Chem., 1992.267.8936.*
- *(4)* P.R. Ortiz **de** Montellano, Y.S. Choe, G. De Pillis, C.E. Catalan0 J. Biol. *Chem.,* **1987.262,** 11641.
- (5) J.A Fruetel, J.R. Collins, D.L. Camper, G.H. Loew, P.R. Ortiz de Montellano J. *Amer. Chem. Sot., 1992,114.6987.*
- *(6)* **D.** Wistuba, H.P. Nowomy, 0. Ttiger, V. **Schurig** *Chirality.* **1989. I.** 127.
- (7) SK Yang. *Biochem. Pharmacol. 1988,37,61.*
- *(8) S.* Colonna, N. Gaggero, A. Manfredi, L. Casella, M. Gullotti, G.Carrea, P. Pasta *Biochemistry,* 1990, 29, 10465; S. Colonna, N. Gaggero, L. Casella, G. Carrea, P. Pasta Tetrahedron Asymmetry, 1992, 3, 95.
- *(9)* D. Mansuy. J. L.eclaire, M. Fontecave, M. Momentau *Biochem. Biophys. Res. Commun.,* **1984,119,** 319.
- (10) J.T. Groves, R.S. Myers J. Amer. Chem. Soc., 1983, 105, 5791.
- (11) C. Querci, M. Ricci J. Chem. Soc. Chem. Commun., 1989, 889.
- (12) R.W. Strassburg, R.A. Gregg, C. Walling J. *Amer. Chem. Sot.,* **1947,69,2 141.**
- **(13) S. Osaki. H. Mimura, N. Yasuhara.** M. Masui, Y. Yamagata, **K.Tomita, T.J. Collins J. Chem. Sot.** *Perkin Trans II,* **1990,353.**